

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/338920913>

An Introduction to Abiotic Stress in Plants

Chapter · January 2020

DOI: 10.22271/ed.book.725

CITATIONS

0

READS

3,231

3 authors:



Jagadish Jena

Indira Gandhi Agricultural University

33 PUBLICATIONS 88 CITATIONS

SEE PROFILE



Soumya Kumar Sahoo

Indira Gandhi Agricultural University

26 PUBLICATIONS 32 CITATIONS

SEE PROFILE



Goutam kumar Dash

ICAR-National Rice Research Institute

55 PUBLICATIONS 135 CITATIONS

SEE PROFILE

Some of the authors of this publication are also working on these related projects:



College of Agriculture (Orissa University of Agriculture & Technology) Chiplima [View project](#)



Drought View project

ADVANCES IN AGRONOMY

VOLUME - 9

Chief Editor

Dr. Anay Kumar Rawat

Assistant Professor

JNKVV, Jabalpur, Madhya Pradesh, India

Co-editor

Uttam Kumar Tripathi

Scientist

J.N.K.V.V, KVK, Chhatarpur, Madhya Pradesh, India

AkiNik Publications

New Delhi

Published By: AkiNik Publications

AkiNik Publications

169, C-11, Sector - 3,

Rohini, Delhi-110085, India

Toll Free (India) – 18001234070

Phone No. – 9711224068, 9911215212

Email – akinikbooks@gmail.com

Chief Editor: Dr. Anay Kumar Rawat

The author/publisher has attempted to trace and acknowledge the materials reproduced in this publication and apologize if permission and acknowledgements to publish in this form have not been given. If any material has not been acknowledged please write and let us know so that we may rectify it.

© **AkiNik Publications**

Publication Year: 2020

Pages: 186

Paperback ISBN: 978-93-89680-59-1

E-Book ISBN: 978-93-89680-60-7

Book DOI: <https://doi.org/10.22271/ed.book.725>

Price: ₹ 576/-

Contents

Chapters	Page No.
1. Laser Controlled Precision Land Levelling <i>(K.K.K. Reddy and G. Vinay)</i>	01-15
2. Technologies for Rice Cultivation in India <i>(Aditya Kumar Singh and Dr. Narendra Singh)</i>	17-42
3. Pulse: Super Food for Sustainable Future <i>(Mousumi Malo and Argha Ghosh)</i>	43-61
4. Review on Changing Scenario of Technological Interventions under Organic Farming in India <i>(Shivam Singh, Jagannath Pathak, Mahendra Pratap Singh, Rudra Pratap Singh and Abhishek Kr. Singh)</i>	63-80
5. Interfacing Crop Simulation Models with GIS for Yield Estimation and Precision Agriculture <i>(Dr. Baby Akula)</i>	81-94
6. Production and Nutritional Quality Enhancement of Nutria-Cereals through Agronomic Manipulations <i>(Mallikarjun Yalagi, Sharanappa and Sahebagouda)</i>	95-118
7. Versatility of Calcium as a Plant Nutrient <i>(Giffy Thomas and Adarsh S)</i>	119-141
8. Waste to Wealth: Recycling of Crop Waste Need for Hour <i>(Sahebagouda, Iranna, Sharanappa and Mallikarjun Yalagi)</i>	143-161
9. An Introduction to Abiotic Stress in Plants <i>(Jagadish Jena, Soumya Kumar Sahoo and Goutam Kumar Dash)</i>	163-186

Chapter - 9

An Introduction to Abiotic Stress in Plants

Authors

Jagadish Jena

Ph.D. Scholar, Department of Agronomy, College of Agriculture, Indira Gandhi Krishi Vishwavidyalaya, Raipur, Chhattisgarh, India

Soumya Kumar Sahoo

Ph.D. Scholar, Department of Plant Physiology, College of Agriculture, Indira Gandhi Krishi Vishwavidyalaya, Raipur, Chhattisgarh, India

Goutam Kumar Dash

Post-Doctoral Fellow, Crop Physiology and Biochemistry Division, ICAR-National Rice Research Institute, Cuttack, Odisha, India

Chapter - 9

An Introduction to Abiotic Stress in Plants

Jagadish Jena, Soumya Kumar Sahoo and Goutam Kumar Dash

Abstract

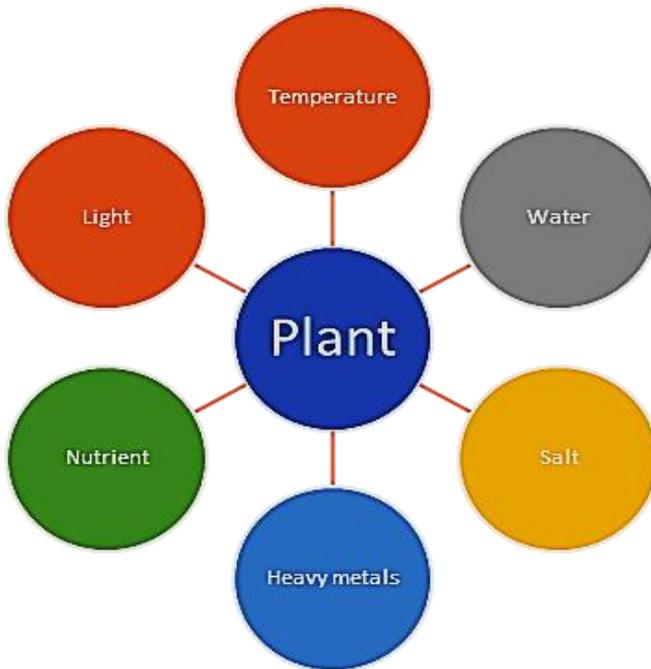
Biological system is an endogenic and dynamic system which always seeks constant in-flow of energy to maintain its meta-stable condition called homeostasis. Any environmental disruption of this homeostasis may be explicated as biological stress. Biological stress limits the growth and yield potential of a crop and has been broadly grouped into biotic and abiotic stresses. Biotic stress conceives due to interactions between organisms starting from predation to allelopathy. Abiotic stress in the other hand appears due to the adverse effects of non-living environmental factors i.e. water, temperature, light, metal, mineral nutrients etc. on plants which are often sporadic and are highly localized. Both of these stresses are inevitable but can be minimized to some extent for increasing the productive potential of crops. Among the management practices for successful crop production in stress condition; breeding, biotechnology and agronomic approaches are widely used. In this chapter an attempt has been made to lucidly present the basic ideas about different types of stress and their impacts on crop production as well as some management approaches to minimize the effects of stress on crops.

Keywords: stress, abiotic stress, temperature stress, heavy metal stress, moisture stress, salinity stress, oxidative stress

Introduction

Climate change is a global issue pertaining abiotic and biotic stress on biosphere directly or indirectly reducing their production and productivity. The concept of stress first developed by Hans Selye in 1936 as unfavorable environmental limitations for plants (Selye H., 1936). Physically, stress is the force per unit area which causes dimensional change on which the stress applied. Biologically, stress is the factors that reduces normal function of individual which limits their genetic potential for growth, development and reproduction (Levitt, 1980). In agricultural context, stress is the factor that

reduces crop productivity and destroys biomass (Grime, 1979). According to Levitt, stress is any environmental factor potentially unfavorable to living organisms (Levitt, 1980). Lichtenthaler extended the concept of plant stress including regeneration phase of plant, differentiating eustress and di-stress (Lichtenthaler, 1988 & 1996). Based on involvement of factors that causes stress, stress can be broadly divided into biotic stress which is the function of interaction between organisms for resources, predation and allelopathic effects and abiotic stress which is the interaction between organism and physical environment. Abiotic factors limit choice of crops, production in large areas and extreme events lead to total crop failure.

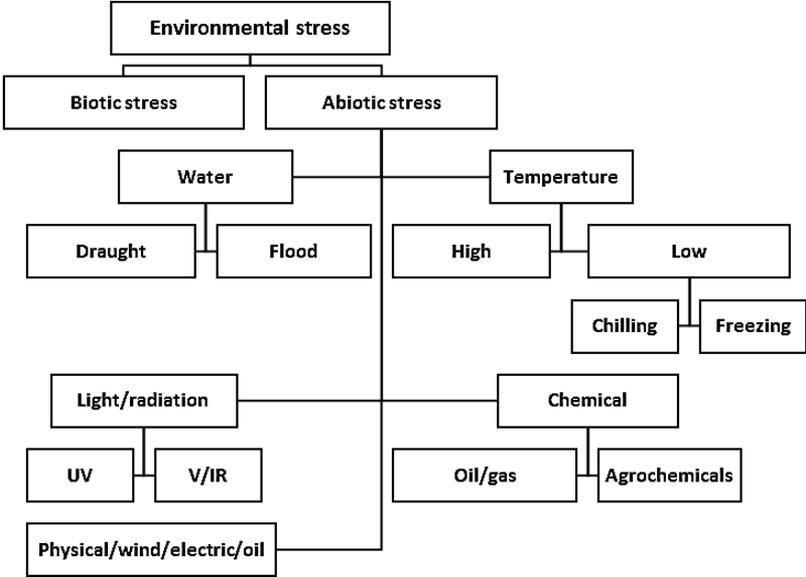


Global climate abnormalities due to different anthropogenic and natural activities typically increased different biotic and abiotic stress on crop plants which affect plants genetic potential of growth and economic production adversely (Prasad *et al.*, 2011; Mahalingam, 2015; Ramegowda and Senthil-Kumar, 2015). Abiotic stress i.e. moisture stress, temperature stress, salinity stress etc. influence occurrence of different biotic stress i.e. pathogens, insects and weeds (Ziska *et al.*, 2010; Peters *et al.*, 2014) at different crop growth stages (Mittler, 2006; Prasad *et al.*, 2011). Abiotic stress may result minor pest to become a potential threat (Duveiller *et al.*, 2007) and may alter plant physiology and defense responses (Schermer and Coakley, 2003). There

are different types of interactions exist between different stress mechanisms and the effect of these interactions on plant growth and productivity may be better understood from “stress matrix” developed by Mittler and colleagues (Mittler, 2006; Suzuki *et al.*, 2014).

Concepts and types of stress

Stress is the factor that exert disadvantageous impacts on plant resulting in unfavorable consequences. This factor may by abiotic i.e. climate and soil or biotic i.e. weeds, pathogens, insects etc. Different types of stress may have different duration for its impact i.e. air temperature can become stressful in few minutes but soil mineral deficiency can take months to become stressful. Stress is measured in terms of plants survival, growth and yield or primary assimilation process i.e. photosynthetic rate, nutrient accumulation, growth rate etc. Stress may be *elastic* when plant recovers with the withdrawal of the stress factor or *plastic* when plants deformed and the change is irreversible after implication of the stress factor.



- **Draught:** It is the stress caused by combination of physical factors reducing water (low moisture stress) availability to the plant sufficient enough to retard the growth and development
- **Flood:** It is the stress caused by excess moisture in the root zone that retard gas exchange (mainly O₂) between soil environment and plant root causing hypoxic condition

- **Heat injury:** Stress caused by temperature more than threshold that can be tolerated by plant i.e. mostly more than 40 °C. When high temperature crosses thermal death point, it leads to death of the plant
- **Freezing/frost injury:** Stress caused by sub-zero temperature i.e. 0 °C or 0 to-10 °C when most of the actively grown plants get killed or injured
- **Chilling injury:** Stress caused by low temperature i.e. 10 to 15 °C or less but more than 0 °C that cause injury mostly in warm season crops i.e. rice, maize, sorghum etc.
- **Radiation injury:** Stress caused by electro-magnetic radiations i.e. UV radiation causing mutation or degradation of biomolecules or IR radiation causing heat injury in plants
- **Salt stress:** Stress caused due to excess soluble salt accumulation in root zone causing specific ion effect or disturbance in mineral nutrient uptake or increasing osmotic potential

Temperature stress

Tropical climate is cursed with higher temperature and radiation which limits the plant growth and development. High temperature causes scorching, sunburn and discolouration of leaves which reduces growth of plants (Vollenweider and Gunthardt-Goerge, 2005). Stress that limits growth, metabolism, and yield potential due to exposure of temperature below or above the thermal threshold for optimal biochemical, physiological and morphological development is called temperature stress (Greaves, 1996). Plants are classified as psychrophiles, mesophiles and thermophiles according to their tolerance to low, medium and high temperature (Levitt, 1980). Adversity of heat stress varies with duration, stage and intensity of the stress (Fahad *et al.*, 2016b). Increase in heat stress reduces numbers of spikelets, numbers of florets per plant in rice, seed set in sorghum (Prasad *et al.* 2006; Fahad *et al.*, 2016b). It has also adverse effects on net assimilation rate and growth in maize and sugarcane and it also reduces nodal length, biomass accumulation, early leaf senescence in sugarcane (Ashraf & Hafeez, 2004; Wahid *et al.*, 2007). Heat stress reduces yield about 50%, 31% and 42% in rice, wheat and maize respectively (Li *et al.*, 2010; Balla *et al.*, 2011; Badu-Apaku *et al.*, 1983). It also reduces the quality due to reduced production of oil, starch and protein (Wilhelm *et al.*, 1999; Maestri *et al.*, 2002).

With the severity of heat stress growth, mass and number of roots reduces significantly due to reduction of water and nutrient supply (Wahid *et*

al., 2007; Hall *et al.*, 2012). Heat stress under limited water supply is proved to be fatal (Machado and Paulsen, 2001). It deactivates enzyme used in chlorophyll biosynthesis (Dutta *et al.*, 2009) which increases the degradation of chlorophyll a & b in developed leaves (Karim *et al.*, 1999). This degradation also associated with oxidative damage. Despite all the physiological biochemical impacts of increased heat stress, the optimum temperature for photosynthetic activity expected to increase with elevated concentration of CO₂ in atmosphere. The injury process in high temperature stress progresses from direct reversible strain i.e. respiration more than photosynthesis to indirect strain i.e. draught or direct strain i.e. starvation (Levitt, 1980). Heat stress tolerant plants show minimal damage to photosynthetic organs, process and increase in biosynthesis of protective compounds (Bita and Gerats, 2013) and the ideal plant types for better tolerance to heat stress should have better photosynthetic rate, membrane thermo-stability and fruit setting and fruit setting under high temperature (Nagarajan *et al.*, 2010).

Heat stress can be managed by developing heat tolerant cultivars through conventional breeding (Ehlers and Hall, 1998), QTLs for grain filling and leaf senescence, heat tolerance during reproductive stage and stay green properties Farooq *et al.*, 2011; Kumar *et al.*, 2010) and their application in transgenic development (Bohnert *et al.*, 2006). Some other heat stress management approaches include pre-conditioning of plants (Morales *et al.*, 2003), pre-sowing or early sowing (Tikhomirova, 1985), application of Ca (Jiang and Huang, 2001), application of Glycine and Betain (Wahid and Shabir, 2005) and application of spermidine (Murkowski, 2001).

Low temperature stress cause wilting, bleaching, browning, necrosis and plant death (Levitt, 1980; Witt and Barfield, 1982). About 15% arable land estimated to be affected by freezing stress (Dudal, 1976).

Moisture stress

Around 28% of world's land are too dry for agricultural support (Kramer and Bouyer, 1995). Estimated annual yield loss due to draught in tropics is nearly 17% (Edmeades *et al.*, 1992). Increased draught with the changing climate scenario results in decrease in physiology, plant growth and reproduction (Barnabas *et al.*, 2008). Daryanto *et al* (2016) reported 21% and 40% yield reduction in wheat and maize respectively due to draught. Draught causes increase in transpiration and reduce water availability to plants roots (Anjum *et al.*, 2011) which tends water balance in negative side affecting growth, nutrient and water relation, photosynthesis

and assimilate partitioning and ultimately the yield (Farooq *et al.*, 2009). Drought stress response in plants varies among species depending on their stages and other growth factors (Demirevska *et al.*, 2010). High temperature stress affect enzyme activity, cell division in plants (Smertenko, 1997) and also changes growing period and distribution of agricultural crops (Porter, 2005).

Drought reduces germination and seedling growth (Kaya *et al.*, 2006; Farooq *et al.*, 2009), reduction in early seedling growth, length of hypocotyl, root and shoot dry weight, vegetative growth in field crops like pea, alfalfa, rice (Manikavelu *et al.*, 2006; Zeid and Shedeed, 2006). Poor crop growth during draught stress mainly due to reduction in cell turgidity (Taiz and Zeiger, 2006) and reduction in mitosis and cell elongation (Hussain *et al.*, 2008). Decreased turgidity reduce leaf expansion causing reduction in photosynthesis and plant growth (Rucker *et al.*, 1995). Pre-anthesis occurrence of draught stress causes early anthesis, while post-anthesis stress causes early grain filling in cereals (Estrada-Campuzano *et al.*, 2008). The decrease in activity of enzymes i.e. sucrose synthase, starch synthase, starch branching enzyme and adenosine di-phosphate glucose pyrophosphorylase which controls grain filling in cereals (Taiz and Zeiger, 2006) is the main cause of yield reduction in cereals (Ahmadi and Baker, 2001). Reduction in rate of photosynthesis (Bota *et al.*, 2004), change in assimilate partitioning (Farooq *et al.*, 2009b) and reduction in flag leaf development (Rucker *et al.*, 1995) are the main causes of yield reduction due to draught stress. Yield loss of 53-92%, 57%, 63-87 reported in rice, wheat and maize respectively due to draught stress (Lafitte *et al.*, 2007; Balla *et al.*, 2011).

Drought stress management can be done by conventional breeding of stress tolerant plant with cultivar having good agronomic performance (Ashraf, 2010), QTL mapping followed by MAS approach (Ashraf *et al.*, 2008), transgenic approach (Ashraf, 2010) and induced stress tolerance i.e. priming (Farooq *et al.*, 2008; Bajwa and Farooq, 2016).

Heavy metal stress

Heavy metals (53 nos.) are those having specific gravity more than 5 g cm⁻³ or atomic mass over 20 and are generally toxic at even low concentration (Sharma and Dietz, 2006; Rascio and Navari-Izzo, 2011) i.e. Cadmium (Cd), Lead (Pb), Arsenic (As), Silver (Ag) etc. Heavy metal contamination in soil are mainly due to human activities i.e. mining, smelting, intensive agricultural practices, fuel production, electroplating etc. (Dembitsky, 2003; Igwe and Abia, 2006; Ali *et al.*, 2013) and may also be

due to natural processes i.e. soil erosion, excessive weathering of rocks and minerals and volcanic eruption. Among heavy metals, some have known physiological functions in plant system called nonessential heavy metals i.e. Arsenic (As), Lead (Pb), Cadmium (Cd), Mercury (Hg) and Selenium (Se) and some are involved in different plant physiological functions like co-factor for enzymatic reactions (Mildvan, 1970) or role in redox reactions (Yruela, 2009) called essential heavy metals i.e. Cobalt (Co), Copper (Cu), Manganese (Mn), Zinc (Zn), Iron (Fe), Molybdenum (Mo) and Nickel (Ni).

Zn accumulation become phytotoxic and inhibits growth by root thickening, reducing cell division and elongation, inhibiting radicle emergence when accumulated at an early stage (Rout and Das, 2009). Increased Zn concentration resulted in disintegration of cell organelles and disruption of membrane in Pigeon pea (Sresty and Rao, 1999), distorted growth in root and leaf of wheat (Pearson and Rengel, 1995) and Mediterranean seagrass (Malea *et al.*, 1995). Ni accumulation above optimal concentration have lethal effect on crop lifecycle, disturbs ionic balance in plant organs (Seregin *et al.*, 2006), show Mg and Fe chlorosis (Piccini and Malavolta, 1992), induce moisture content followed by disturbing stomatal conductance and photosynthetic activity in cabbage (Molas, 1997) and disturb plant nutrient homeostasis and equilibrium (Rubio *et al.*, 1994). Cd is a nonessential heavy metal and its hyper accumulation causes stunted growth and chlorosis in plants because of reduced Fe uptake (Haghiri, 1974), interfere transport and uptake essential nutrients i.e. P, K, Ca, Mg, Mn etc. and plant water uptake (Godbold and Huttermann, 1985).

The removal of these heavy metals from root zone of soil may done economically and efficiently using phytoremediation techniques (Salt *et al.*, 1995) that includes phytoextraction, phytosequestration, phytodegradation, phytovolatilization, phytohydraulics and rhizodegradation (Tsao, 2003). Phytoextraction one of the attractive tools among these phytoremediation techniques which uses hyperaccumulators i.e. species from *Arabidopsis*, *Thalaspia* and *Pteris* genus (Hanikenne *et al.*, 2008 and Blande *et al.*, 2017) which can accumulate extremely high levels of heavy metals in their aerial parts without any toxic sign (Rascio and Navari-Izzo, 2011). Generally, plants containing higher than 0.1 mg g⁻¹ of Cd, 1 mg g⁻¹ of As, Cu, Cr, Co, Pb, Ni or 10 mg g⁻¹ of Mg or Zn in their dry matter are considered to be as metal hyperaccumulators (Salt *et al.*, 1998). There are certain mechanisms for heavy metal tolerance in hyperaccumulators i.e. enhanced uptake in root symplasm, efficient xylem loading for root to shoot transport, complex formation with nicotinamine for transport, accumulation mainly in aerial

organs, complex formation with weak ligands such as malate or citrate for storage, sequester generally in vacuole of epidermal cells and strongly enhanced anti-oxidants whereas, heavy metal tolerance mechanism in non-hyperaccumulators are i.e. metals mostly accumulated in roots, small amount translocated into shoots, small portion accumulated in epidermal and mesophyll cells of shoots, form complexes with strong ligands such as PCs, glutathione for storage and enhanced anti-oxidants. There are also certain gene families or biosynthesis pathways with respect to adaptation mechanisms in plants under heavy metal stress conditions during phytoremediation which can be used for transgenic plant development which may be capable of remediating heavy metals in soil.

Salt/salinity stress

Crop said to be in salt stress when it is unable to express its full genetic potential in terms of growth, development and reproduction as the salinity in the soil exceeds the critical level (Grieve *et al.*, 2012). The composition and concentration of dissolved salts in soil and irrigation water varies from one place to another (Tanji and Wallender, 2012). The adverse effect of salt affected soils may be due to high salt concentration in the soil solution i.e. osmotic effects or by high concentration of specific ions such as sodium or chloride that can cause injury to sensitive crops i.e. specific ion effect. Adverse effect of saline soil is due to soluble salt concentration whereas, adverse effects of sodic soil is due to deterioration of soil physical condition (Shainberg and Singer, 2012).

The adverse effect of salt stress may be due to specific ion effect i.e. Na⁺ and Cl⁻ (Kingsbury and Epstein, 1986; Munns and Termaat, 1986) or interacting with other mineral nutrient dynamics (Shabala and Munns, 2012).

Most of the plants are non-halophytes or glycophytes which hardly tolerate high salt concentration i.e. rice, wheat, peas etc. and some plants are halophytes in nature which can tolerate high salt concentration and are from native flora of extremely saline soils i.e. mangroves, *Atriplex* spp., *Salicornia* spp. etc. Most of the osmotic adjustments can be achieved due to either the absorption or accumulation of ions from the medium or by synthesis and accumulating organic solutes and their dominance dependent on the type of plants and the level of salinity. Halophytes are generally adapted to high saline environment by accumulating excess absorbing salts in osmoticum in vacuole (Flowers and Colmer, 2008) or osmotic adjustment by different organic solutes in the cytoplasm (Wyn Jones and Gorham, 2002), sequestering excess salt from metabolically active tissues in salt

glands and salt bladders (Flowers and Colmer, 2008; Shabala and Munns, 2012), secretion of compatible solutes or osmolytes i.e. proline, glycine betain and sucrose (Munns and Tester, 2008), osmoprotectant nature of these organic solutes (McNeil *et al.*, 1999), ion selective absorption and compartmentalization (Flowers, 2008), scavenging of reactive oxygen species (Viswanathan *et al.*, 2001) and salt tolerant gene expression (Byrt *et al.*, 2014).

Nutrient stress

There are several mineral elements contributing to the growth and development of a plant, among which 17 are called essential nutrients according to the essentiality criteria defined by Arnon and Stout. Mineral nutrition being an independent discipline of plant physiology (Grossman and Takahashi, 2001), essential minerals are further divided into four groups by Mengel and Kirkby (1978) according to their biological structures and metabolic functions. There is certain nutrient stress (deficiency or excess) reported by several scientists in different plants. Nitrate may be involved in biosynthesis and transport of cytokinin (Liu *et al.*, 2000) and higher level of nitrate (NO_3^-) inhibits root growth and root: shoot ratio (Zhang *et al.*, 1999). Phosphorous deficiency limited primary root elongation and enhance lateral root formation (Hodge, 2004), decrease shoot-root dry weight ratio (Fredeen *et al.*, 1989), reduce leaf number (Lynch *et al.*, 1991), affect formation of reproductive organs (Barry and Miller, 1989), decreased photosynthetic rate in soybean (Lauer *et al.*, 1989) and induced the expression of phosphoenolpyruvate carboxylase (PEPCO) in tobacco (Toyota *et al.*, 2003). Potassium (K^+) deficient plants are susceptible to lodging and draught (Lindhauer, 1985). Sulfur deficiency decreased net photosynthesis and root hydraulic conductivity (Karmoker *et al.*, 1991), decrease in shoot-root dry weight ratio (Edelbauer, 1980), impaired carbohydrate metabolism followed by induced starch accumulation (Willenbrink, 1967). Calcium deficiency cause premature fruit and bud shedding and excess calcium interferes magnesium absorption. Magnesium deficiency cause superoxide free radical accumulation (Cakmak and Marschner, 1992). Iron deficient plants are low in chlorophyll content, inhibit photosynthetic electron transport with reduced RUBP regeneration, responsible for low starch and sugar content and low CO_2 fixation (Sharma and Sanwal, 1992). Iron toxicity in water logged soil may cause bronzing, stunted top and root growth. Siderophores are the organic molecules which combine with iron and make it available to plants and microbes (Brown *et al.*, 1991). Manganese deficiency caused reduced root growth (Marcar and Graham, 1987) and some other typical deficiency symptoms of manganese deficiency in plants are *grey speck* of oats, *marsh*

spot of pea and *Pahala blight* of sugarcane. Zinc deficiency causes decrease in protein content may be attributed due to unstable ribosomes (Obata and Umbebayashi, 1988) and low level of RNA (Cakmak *et al.*, 1989) in low zinc availability. Copper deficiency in the leaves of subterranean clover reduced activity of polyphenol oxidase (Delhaize *et al.*, 1985). Copper deficiency also decreased activity of IAA oxidase and peroxidase (Davies *et al.*, 1978). It also causes die back in citrus, interveinal chlorosis and distorted lamina in legumes and tomato. Molybdenum deficiency in mustard leaves showed reduced level of DNA and RNA (Chatterjee *et al.*, 1985). Molybdenum deficient plants are less resistant to low temperature and water logging stress (Vunkova-Radeva *et al.*, 1988). The common boron deficiency symptoms are tobacco “top sickness”, sugar beet “heart rot”, celery “stem crack” and cauliflower “hollow heart”. Boron deficiency inhibit DNA synthesis (Krueger *et al.*, 1987), depressed cytokinin synthesis in sunflower (Wagner and Michael, 1971) and decrease RNA content due to enhanced RNase activity (Dave and Kannan, 1980).

Oxidative stress

Molecular oxygen (O_2) in atmosphere are results of photosynthesis and are required for basic aerobic life. O_2 fixed in various biomolecules by different non-enzymatic and enzymatic processes (Gilbert, 1981; Elstner, 1982, 1987). Ground state dioxygen (O_2) is unreactive for biomolecules but its physical activation i.e. singlet oxygen or chemical activation i.e. superoxide (O_2^-), hydrogen peroxide (H_2O_2) and hydroxyl radicals (OH) are biologically toxic (McKersie and Leshem, 1994). Superoxide and hydrogen peroxide both can be act as oxidant and reductant but hydroxyl radicals are the most powerful oxidizing agent that don't have any specific scavenger to neutralize it. Different reactive oxygen species (ROS) i.e. O_2^- , H_2O_2 and OH causes bleaching by chlorophyll degradation, lipid peroxidation, inhibition of sensitive enzymes, base alteration of DNA (Kasai *et al.*, 1986), proteolysis (Wolff *et al.*, 1986) etc. Reactive oxygen species are produced in chloroplast (Asada, 2006), mitochondria, endoplasmic reticulum, peroxisomes and glyoxysomes, plasma membrane and the apoplast compartment (McKersie and Leshem, 1994).

ROS in plants generally formed whenever they are subjected to different stress i.e. photoinhibition (Sonoike, 1996), herbicide, toxins and triazole compounds (Kauss and Jeblick, 1995), heavy metal toxicity (Foyer *et al.*, 1994), UV radiation (Hideg and Vass, 1996), salt stress (Miszalski *et al.*, 1998), draught and heat stress (Filek *et al.*, 1997; Biemelt *et al.*, 1998), chilling and freezing (Tao *et al.*, 1998).

There are several defense systems in plants to protect plants from these ROS i.e. superoxide dismutase (SOD), ascorbate peroxidase, glutathione reductase (GR), mono-dehydroxy ascorbate reductase, catalase (CATs). Except these enzymatic antioxidants, there are some non-enzymatic antioxidants which play important role to protect biomolecules from ROS i.e. ascorbate, tocopherol, carotenoids, glutathione etc. Among all these antioxidants, SOD gives primary defense against oxygen free radicals (Bannister *et al.*, 1987) converting O_2^- to H_2O_2 and protects plants from different stress (Mittler and Zilinskas, 1994; Allen *et al.*, 1997).

References

1. Ahmadi A, Baker DA. The effect of water stress on the activities of key regulatory enzymes of the sucrose to starch pathway in wheat, *Plant Growth Regul.* 2001; 35:81-91.
2. Ahmadi AA. Effect of post-anthesis water stress on yield regulating processes in wheat (*Triticum aestivum* L.). Ph.D. Thesis. University of London, Wye College, Wye, Ashford, U.K, 1998.
3. Ali H, Khan E, Sajad MA. Phytoremediation of heavy metals-concepts and applications. *Chemosphere.* 2013; 91(91):869-881.
4. Allen RD, Webb RP, Schake SA. Use of transgenic plants to study antioxidant defenses. *Free Rad. Biol. Med.* 1997; 23:473-479.
5. Anjum SA, Xie XYU, Wang L, Chang Saleem MF, Man C, Lei W. Morphological, physiological and biochemical responses of plants to drought stress. *African Journal of Agricultural Research.* 2011; 6:2026-2032.
6. Asada K. Production and action of active oxygen species in photosynthetic tissues. In Foyer CH, Mullineaux PM (eds.), *Causes of Photooxidative Stress and Amelioration of Defense Systems in Plants*, CRC Press, Boca Raton, 1994, 77-104.
7. Ashraf M. Inducing drought tolerance in plants: recent advances. *Biotechnol. Adv.* 2010; 28:169-183.
8. Ashraf M, Hafeez M. Thermotolerance of pearl millet and maize at early growth stages: Growth and nutrient relations. *Biol. Plant.* 2004; 48(1):81-86.
9. Ashraf M, Athar HR, Harris PJC, Kwon TR. Some prospective strategies for improving crop salt tolerance. *Adv. Agron.* 2008; 97:45-110.

10. Bajwa AA, Farooq M. Seed priming with sorghum water extract beneficial bacteria of agricultural importance. *Biotechnol. Lett.* 32, 1559-1570. Surfactant improves germination metabolism and early seedling growth of wheat. *Arch. Agron. Soil Sci.* 2016; 63:319-329.
11. Balla K, Rakszegi M, Li Z, Békés F, Bencze S, Veisz O. Quality of winter wheat in relation to heat and drought shock after anthesis. *Czech J Food Sci.* 2011; 29(2):117-128.
12. Balla K, Rakszegi M, Li Z, Békés F, Bencze S, Veisz O. Quality of winter wheat in relation to heat and drought shock after anthesis. *Czech J Food Sci.* 2011; 29(2):117-128.
13. Bannister JV, Bannister WH, Rotilio G. Aspects of the structure, function, and applications of superoxide dismutase. *CRC Crit. Rev. Biochem.* 1987; 22:111-180.
14. Barnabas B, Jager K, Feher A. The effect of drought and heat stress on reproductive processes in cereals. *Plant, Cell & Environment.* 2008; 31:11-38.
15. Barry DAJ, Miller MH. Phosphorus nutritional requirement of maize seedlings for maximum yield. *Agron. J.* 1989; 81:95-99.
16. Biemelt S, Keetman U, Albrecht G. Re-aeration following hypoxia or anoxia leads to activation of the antioxidative defense system in roots of wheat seedlings. *Plant Physiol.* 1998; 116:651-658.
17. Bitá CE, Gerats T. Plant tolerance to high temperature in a changing environment: Scientific fundamentals and production of heat stress-tolerant crops. *Front. Plant Sci.* 2013; 4(JUL):1-18.
18. Blande D, Halimaa P, Tervahauta AI, Aarts MGM, Kärenlampi SO. De novo transcriptome assemblies of four accessions of the metal hyperaccumulator plant *Noccaea caerulescens*. *Sci. Data*, 2016, 1-9.
19. Bohnert HJ, Gong Q, Li P, Ma S. Unraveling abiotic stress tolerance mechanisms-Getting genomics going. *Curr. Opin. Plant Biol.* 2006; 9(2):180-188.
20. Bota J, Flexas J, Medrano H. Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress? *New Phytol.* 2004; 162:671-681.
21. Brown JC, Jolley VD, Lytle CM. Comparative evaluation of iron solubilizing substances (phytosiderophores) released by oats and corn: Iron-efficient and iron-inefficient plants. *Plant Soil.* 1991; 130:157-163.

22. Byrt CS, Xu B, Krishnan M, Lightfoot DJ, Athman A, Jacobs AK *et al.* The Na⁺ transporter, TaHKT1; 5-D, limits shoot Na⁺ accumulation in bread wheat. *Plant Journal*. 2014; 80:516-526.
23. Cakmak I, Marschner H. Magnesium deficiency and high light intensity enhance activities of superoxide dismutase, ascorbate peroxidase and glutathione reductase in bean leaves. *Plant Physiol*. 1992; 98:1222-1227.
24. Cakmak I, Marschner H, Bangerth F. Effect of zinc nutritional status on growth, protein metabolism and levels of indole-3 acetic acid and other phytohormones in bean (*Phaseolus vulgaris* L.). *J Exp. Bot.* 1989; 40:404-412.
25. Chatterjee C, Nautiyal N, Agarwala SC. Metabolic changes in mustard plant associated with molybdenum deficiency. *New Phytol.* 1985; 100:511-518.
26. Daryanto S, Wang L, Jacinthe PA. Global synthesis of drought effects on maize and wheat production. *PLOS One*. 2016; 11(5):1-15.
27. Dave IC, Kannan S. Boron deficiency and its associated enhancement of RNase activity in bean plants. *Z. Pflanzenphysiol.* 1980; 97:261-264.
28. Davies JN, Adams P, Winsor GW. Bud development and flowering of *Chrysanthemum morifolium* in relation to some enzyme activities and to the copper, iron and manganese status. *Commun. Soil Sci. Plant Anal.* 1978; 9:249-264.
29. Delhaize E, Loneragan JF, Webb J. Development of three copper metalloenzymes in clover leaves. *Plant Physiol*, 1985, 78:4-7.
30. Dembitsky V. Natural occurrence of arseno compounds in plants, lichens, fungi, algal species, and microorganisms. *Plant Sci.* 2003; 165:1177-1192.
31. Demirevska K, Simova-Stoilova L, Fedina I, Georgieva K, Kunert K. Response of oryzacystatin I transformed tobacco plants to drought, heat and light stress. *Journal of Agronomy and Crop Science*. 2010; 196:90-99.
32. Dudal R. Inventory of major soils of the world with special reference to mineral stress.-*Plant Adaption to Mineral Stress in Problem Soils*. Ed. M. J Wright. Cornell Univ. Agric. Exp. Stn. Ithaca, N.Y, 1976, 3-23.
33. Dutta S, Mohanty S, Tripathy BC. Role of temperature stress on chloroplast biogenesis and protein import in pea [OA]. *Plant Physiology*. 2009; 150(2):1050-1061.

34. Duveiller E, Singh RP, Nicol JM. The challenges of maintaining wheat productivity: pests, diseases and potential epidemics. *Euphytica*. 2007; 157:417-430.
35. Edelbauer A. Auswirkung von abgestuftem Schwefelmangel auf Wachstum, Substanzbildung und Mineralstoffgehalt von Tomate (*Lycopersicon esculentum* Mill.) In: Nährlosungskultur. Die Bodenkultur. 1980; 31:229-241.
36. Edmeades GOJ, Bolaoos, Lafitte HR. Progress in selecting for drought tolerance in maize. In D. Wilkinson (ed.), Proc. 47th Annual Corn and Sorghum Research Conference, Chicago, December 9ñ10. ASTA, Washington, 1992, 93n111.
37. Ehlers JD, Hall AE. Heat tolerance of contrasting cowpea lines in short and long days. *F. Crop. Res.* 1998; 55(1-2):11-21.
38. Elstner EF. Oxygen activation and oxygen toxicity. *Annu. Rev. Plant Physiol.* 1982 33:73-96.
39. Estrada-Campuzano G, Miralles DJ, Slafer GA. Genotypic variability and response to water stress of pre- and post-anthesis phases in triticale. *Eur. J Agron.* 2008; 28(3):171-177.
40. Fahad S, Hussain S, Saud S, Khan F, Hassan S, Amanullah *et al.* exogenously applied plant growth regulators affect heat-stressed rice pollens. *J Agron. Crop Sci.* 2016b; 202:139-150.
41. Farooq M, Basra SMA, Rehman H, Saleem BA. Seed priming enhances the performance of late sown wheat (*Triticum aestivum* L.) by improving the chilling tolerance. *J Agron Crop Sci.* 2008; 194:55-60.
42. Farooq M, Bramley H, Palta JA, Siddique KHM. Heat stress in wheat during reproductive and grain-filling phases. *CRC. Crit. Rev. Plant Sci.* 2011; 30(6):491-507.
43. Farooq M, Wahid A, Kobayashi N, SMA, Fujita DB. Plant drought stress: effects, mechanisms and management To cite this version: Review article. *Agron. Sustain. Dev.* 2009; 29(1):185-212.
44. Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA. Plant drought stress: effects, mechanisms and management. *Agron. Sustain. Dev.* 2009b; 29:185-212.
45. Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA. Plant drought stress: Effects, mechanisms and management. In *Sustainable Agriculture*, (Springer Netherlands), 2009, 153-188.

46. Filek M, Baczek R, Niewiadomska E, Pilipowicz M, Koscielniak J. Effect of high temperature treatment of *Vicia faba* roots on the oxidative stress enzymes in leaves. *Acta Biochim. Pol.* 1997; 44:315-321.
47. Flowers TJ, Colmer TD. Salinity tolerance in halophytes. *New Phytologist.* 2008; 179:945-963.
48. Flowers TJ, Colmer TD. Salinity tolerance in halophytes. *New Phytologist.* 2008; 179:945-963.
49. Foyer CH, Harbinson J. Oxygen metabolism and the regulation of photosynthetic electron flow. In Foyer CH, Mullineaux PM, (eds.), *Causes of Photooxidative Stress and Amelioration of Defense Systems in Plants*, CRC Press, Boca Raton, 1994, 1-42.
50. Fredeen AL, Rao IM, Terry N. Influence of phosphorus nutrition on growth and carbon partitioning in *Glycine max*. *Plant Physiol.* 1989; 89:225-230.
51. Godbold DL, Huttermann A. Effect of zinc, cadmium and mercury on root elongation of *Picea abies* (Karst.) seedlings, and the significance of these metals to forest die-back. *Environ. Pollu.* 1985; 38:375-381.
52. Greaves J. The gender trap. *Health Informatics J.* 1996; 2(4):194-198.
53. Grieve CM, Grattan SR, Maas EV. *Plant Salt Tolerance. Agricultural Salinity. Assessment and Management* (2nd Edition). 2012; 71:405-459.
54. Grime JP. *Plant strategies and vegetation processes*. Chichester: Wiley and Sons: 222, 1979.
55. Grossman A, Takahashi H. Macronutrient utilization by photosynthetic eukaryotes and the fabric of interactions, *Annu. Rev. Plant Phys.* 2001; 52:163-210.
56. Haghiri F. Plant uptake of cadmium as influenced by cation exchange capacity, organic matter, Zinc, and soil temperature. *J Environ. Qual.* 1974; 3:180-183.
57. Hall AE. Heat stress. *Plant Stress Physiol.* CABI Publishing, 2012, 118-131.
58. Hanikenne M, Talke IN, Haydon MJ, Lanz C, Nolte A, Motte P *et al.* Evolution of metal hyperaccumulation required cis-regulatory changes and triplication of HMA4. *Nature.* 2008; 453(7193):391-395.
59. Hideg E, Vass I. UV-B induced free radical production in plant leaves and isolated thylakoid membranes. *Plant Sci.* 1996; 115:251-260.

60. Hodge A. The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytol.* 2004; 162:9-24.
61. Hussain AI, Anwar F, Hussain Sherazi ST, Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chem.* 2008; 108(3):986-995.
62. Igwe JC, Abia AA. A bioseparation process for removing heavy metals from waste water using biosorbents. *Afr. J Biotechnol.* 2006; 5:1167-1179
63. Jiang Y, Huang B. Drought and heat stress injury to two cool-season turfgrasses in relation to antioxidant metabolism and lipid peroxidation. *Crop Sci.* 2001; 41(2):436-442.
64. Karim MA, Fracheboud Y, Stamp P. Photosynthetic activity of developing leaves may be less affected by heat stress than that of developed leaves. *Physiol. Plant.* 1999; 105:685-693.
65. Karmoker JL, Clarkson DL, Saker LR, Rooney JM, Purves JV. Sulphate deprivation depresses the transport of nitrogen to the xylem and the hydraulic conductivity of barley (*Hordeum vulgare* L.) roots. *Planta.* 1991; 185:269-278.
66. Kasai H, Crain PF, Kuchino Y, Nishimura S, Ootsuyama A, Tanooka H. Formation of 8-hydroxyguanine moiety in cellular DNA by agents producing oxygen radicals and evidence for its repair. *Carcinogenesis.* 1986; 7:1849-1851.
67. Kaus H, Jeblick W. Pretreatment of parsley suspension cultures with salicylic acid enhances spontaneous and elicited production of H₂O₂. *Plant Physiol.* 1995; 108:1171-1178.
68. Kaya C, Tuna L, Higgs D. Effect of silicon on plant growth and mineral nutrition of maize grown under water-stress conditions. *J Plant Nutr.* 2006; 29(8):1469-1480.
69. Kingsbury RW, Epstein E. Salt sensitivity in wheat. A case for specific ion toxicity. *Plant Physiology.* 1986; 80:651-654.
70. Kramer PJ, Boyer JS. *Water relations of plants and soils.* Academic Press, San Diego, 1995.
71. Krueger RW, Lovatt CJ, Albert LS. Metabolic requirement of *Cucurbita pepo* for boron. *Plant Physiol.* 1987; 83:254-258.

72. Kumar U, Joshi AK, Kumari M, Paliwal R, Kumar S, Röder MS. Identification of QTLs for stay green trait in wheat (*Triticum aestivum* L.) in the “Chirya 3” × “Sonalika” population. *Euphytica*. 2010; 174(3):437-445.
73. Lafitte HR, Yongsheng G, Yan S, Li ZK. Whole plant responses, key processes, and adaptation to drought stress: the case of rice, *J Exp. Bot.* 2007; 58:169-175.
74. Lauer MJ, Blevins DG, Sierzputowska-Gracz H. ³¹P-nuclear magnetic resonance determination of phosphate compartmentation in leaves of reproductive soybeans (*Glycine max* L.) as effected by phosphate nutrition. *Plant Physiol.* 1989; 89:1331-1336.
75. Levitt J. Responses of Plants to Environmental Stress. In *Water, Radiation, Salt and other Stress*. New York: New York: Academic Press, 1980.
76. Lichtenthaler HK. Vegetation Stress: an Introduction to the Stress Concept in Plants. *Journal of Plant Physiology.* 1996; 148:4-14.
77. Lichtenthaler HK, Rinderle U. The Role of Chlorophyll Fluorescence in the Detection of Stress Conditions in Plants. *CRC Critical Reviews in Analytical Chemistry.* 1988; 19:S29-S85.
78. Lindhauer MG. Influence of K nutrition and drought on water relation and growth of sunflower (*Helianthus annuus* L.). *Z. Pflanzenernahr. Bodenk.* 1985; 148:654-669.
79. Lynch J, Lauchli A, Epstein E. Vegetative growth of the common bean in response to phosphorus nutrition. *Crop Sci.* 1991; 31:380-387.
80. Machado S, Paulsen GM. Combined effects of drought and high temperature on water relations of wheat and sorghum. *Plant and Soil.* 2001; 233(2):179-187.
81. Maestri E, Klueva NK, Perrotta C, Gulli M, Nguyen H, Marmioli N. Molecular genetics of heat tolerance and heat shock proteins in cereals. *Plant Mol. Biol.* 2002; 48:667-681.
82. Mahalingam R (Ed.). “Consideration of combined stress: a crucial paradigm for improving multiple stress tolerance in plants,” in *Combined Stresses in Plants*, (Cham: Springer International Publishing), 2015, 1-25.
83. Malea P, Kevrekidis T, Haritonidis S. The short-term uptake of zinc and cell mortality of the sea grass *Halophila stipulacea* (Forsk.) Aschers, Israel. *J Plant Sci.* 1995; 43:21-30.

84. Manikavelu A, Nadarajan N, Ganesh SK, Gnanamalar RP, Babu RC. Drought tolerance in rice: morphological and molecular genetic consideration, *Plant Growth Regul.* 2006; 50:121-138.
85. Marcar NE, Graham RD. Genotypic variation for manganese efficiency in wheat. *J Plant Nutr.* 1987; 10:2049-2055.
86. Mckersie BD, Leshem YY. *Stress and Stress Coping in Cultivated Plants.* Kluwer Academic Publishers, Dordrecht, 1994, 1-256.
87. McNeil SD, Nuccio ML, Hanson AD. Betaine and related osmoprotectants. Targets for metabolic engineering of stress resistance. *Plant Physiology.* 1999; 120:945-949.
88. Mengel K, Kirkby EA. *Principles of plant nutrition.* Bern. Int. Potash Inst, 1978, 593.
89. Mildvan AS. Metal in enzymes catalysis. *Enzymes.* 1970; 11(11):445-536.
90. Mittler R. Abiotic stress, the field environment and stress combination. *Trends Plant Sci.* 2006; 11:15-19.
91. Mittler R, Zilinskas BA. Regulation of pea cytosolic ascorbate peroxidase and other antioxidant enzymes during the progression of drought stress and following recovery from drought. *Plant J.* 1994; 5:397-405.
92. Molas J. Changes in morphological and anatomical structure of cabbage (*Brassica oleracea* L.) outer leaves and in ultra-structure of their chloroplasts caused by an *in vitro* excess of nickel. *Photosynthetica.* 1997; 34:513-522.
93. Morales D, Rodríguez P, Dell'Amico J, Nicolás E, Torrecillas A, Sánchez-Blanco MJ. High-temperature preconditioning and thermal shock imposition affects water relations, gas exchange and root hydraulic conductivity in tomato. *Biol. Plant.* 2003; 47(2):203-208.
94. Munns R, Fisher DB, Tonnet ML. Na⁺ and Cl⁻ transport in the phloem from leaves of NaCl-treated barley. *Australian Journal of Plant Physiology.* 1986; 13:757-766.
95. Munns R, Tester M. Mechanisms of salinity tolerance. *Annual Review of Plant Biology.* 2008; 59:651-685.
96. Murkowski A. Heat stress and spermidine: Effect on chlorophyll fluorescence in tomato plants. *Biol. Plant.* 2001; 44(1):53-57.

97. Nagarajan R, Rajmohan N, Mahendran Senthamilkumar U. Evaluation of groundwater quality and its suitability for drinking and agricultural use in Thanjavur city, Tamil Nadu, India, *Environ Monit Assess.* 2010; 171:289-308.
98. Obata H, Umebayashi M. Effect of zinc deficiency on protein synthesis in cultures tobacco plant cells. *Soil Sci. Plant Nutr.* (Tokyo). 1988; 34:351-357.
99. Pearson JN, Rengel Z. Uptake and distribution of ^{65}Zn and ^{54}Mn in wheat grown at sufficient and deficient levels of Zn and Mn: I. During vegetative growth. *J Exp. Bot.* 1995; 46(7):833-839.
100. Peters K, Breitsameter L, Gerowitt B. Impact of climate change on weeds in agriculture: a review. *Agric. Sustain. Dev.* 2014; 34:707-721.
101. Piccini DF, Malavolta E. Effect of nickel on two common bean cultivars. *J Plant Nut.* 1992; 15:2343-2350.
102. Porter JR. Rising temperatures are likely to reduce crop yields. *Nature.* 2005; 436:174.
103. Prasad PVV, Boote KJ, Allen LH. Adverse high temperature effects on pollen viability, seed-set, seed yield and harvest index of grain-sorghum [*Sorghum bicolor* (L.) Moench] are more severe at elevated carbon dioxide due to higher tissue temperatures. *Agric. For. Meteorol.* 2006; 139(3-4):237-251.
104. Prasad PVV, Pisipati SR, Momčilović I, Ristic Z. Independent and Combined Effects of High Temperature and Drought Stress During Grain Filling on Plant Yield and Chloroplast EF-Tu Expression in Spring Wheat. *J Agron. Crop Sci.* 2011; 197(6):430-441.
105. Ramegowda V, Senthil-Kumar M. The interactive effects of simultaneous biotic and abiotic stresses on plants: mechanistic understanding from drought and pathogen combination. *J Plant Physiol.* 2015; 176:47-54.
106. Rascio N, Navari-Izzo F. Heavy metal hyperaccumulating plants: how and why do they do it? And what makes them so interesting? *Plant Sci.* 2011; 180(2):169-181.
107. Rout GR, Das P. Effect of metal toxicity on plant growth and metabolism: Zinc I, Eric L, Navarrete M, Debaeke P, Véronique S, Alberola C (Eds.), *Sustainable agriculture*, Springer Netherlands, 2009, 873-884.

108. Rubio MI, Escrig I, Martínez-Cortina C, López-Benet FJ, Sanz A. Cadmium and nickel accumulation in rice plants. Effects on mineral nutrition and possible interactions of abscisic and gibberellic acids. *Plant Growth Regul.* 1994; 14:151-157.
109. Rucker KS, Kvien CK, Holbrook CC, Hook JE. Identification of peanut genotypes with improved drought avoidance traits. *Peanut Sci.* 1995; 24:14-18.
110. Salt DE, Blaylock M, Kumar NPBA, Viatcheslav D, Ensley BD *et al.* Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. *Bio-Technology.* 1995; 13:468-74.
111. Salt DE, Smith RD, Raskin L. Phytoremediation. *Ann Rev Plant Phys Plant Mol Biol.* 1998; 49(1):643-668.
112. Scherm H, Coakley SM. Plant pathogens in a changing world. *Australas. Plant Pathol.* 2003; 32:157-165.
113. Selye H. *Nature* 36. Pdf. *Nature*, 1946.
114. Seregin IV, Kozhevnikova AD, Kazyumina EM, Ivanov VB. Nickel toxicity and distribution in maize roots. *Fiziol Rast.* 2006; 50:793-800.
115. Shabala S, Munns R. Salinity stress: Physiological constraints and adaptive mechanisms. In: Shabala S. (Ed.), *Plant Stress Physiology.* London, UK: CAB International, 2012.
116. Shainberg I, Singer MJ. Soil response to saline and sodic conditions. In: Wallender WW, Tanji KK (Eds.), *Agricultural Salinity Assessment and Management.* ASCE Manual and Reports on Engineering Practice No. 71, second Ed. New York, NY: American Society of Civil Engineers, 2012, 139-167.
117. Sharma S, Sanwal GG. Effect of Fe deficiency on the photosynthetic system of maize. *J Plant Physiol.* 1992; 140:527-530.
118. Sharma SS, Dietz KJ. The significance of amino acids and amino acid-derived molecules in plant responses and adaptation to heavy metal stress. *J Exp. Bot.* 2006; 57(4):711-726.
119. Smertenko A, Dráber P, Viklický V, Opatrný Z. Heat stress affects the organization of microtubules and cell division in *Nicotiana tabacum* cells. *Plant, Cell Environ.* 1997; 20(12):1534-1542.
120. Sonoike K. Photoinhibition of photosystem I: its physiological significance in the chilling sensitivity of plants. *Plant Cell Physiol.* 1996; 37:239-247.

121. Sresty TVS, Madhava Rao KV. Ultrastructural alterations in response to zinc and nickel stress in the root cells of pigeonpea. *Environ. Exp. Bot.* 1999; 41(1):3-13.
122. Suzuki N, Rivero RM, Shulaev V, Blumwald E, Mittler R. Abiotic and biotic stress combinations. *New Phytol.* 2014; 203:32-43.
123. Taiz L, Zeiger E. *Plant physiology*. 4th Edition, Sinauer Associates, Inc., Sunderland, 2006.
124. Taiz L, Zeiger E. *Plant physiology*. 4th Edition, Sinauer Associates, Inc., Sunderland, 2006.
125. Tanji KK, Wallender WW. Nature and extent of agricultural salinity and sodicity. In: Wallender WW, Tanji KK (Eds.), *Agricultural Salinity Assessment and Management*. ASCE Manuals and Reports on Engineering Practice No. 71, second review New York, NY: American Society of Civil Engineers, 2012, 1-25.
126. Tao DL, Oquist G, Wingsle G. Active oxygen scavengers during cold acclimation of Scots pine seedlings in relation to freezing tolerance. *Cryobiology*. 1998; 37:38-45.
127. Tikhomirova EV. Changes of nitrogen metabolism in millet at elevated temperatures. *Field Crops Research*. 1985; 11(1985):259-264.
128. Toyota K, Koizumi N, Sato F. Transcriptional activation of Phosphoenol pyruvate carboxylase by Phosphorus deficiency in tobacco. *J Exp. Bot.* 2003; 54:961-969.
129. Tsao DY, Freiwald WA, Knutsen TA, Mandeville JB, Tootell RBH. Faces and objects in macaque cerebral cortex. *Nat. Neuro. Sci.* 2003; 6(9):989-995.
130. Viswanathan C, Khanna-Chopra R. Effect of heat stress on grain growth, starch synthesis and protein synthesis in grains of wheat (*Triticum aestivum* L.) varieties differing in grain weight stability. *J Agron. Crop Sci.* 2001; 186:1-7.
131. Vollenweider P, Günthardt-Goerg MS. Diagnosis of abiotic and biotic stress factors using the visible symptoms in foliage. *Environ. Pollut.* 2005; 137(3):455-465.
132. Vunkova-Radeva R, Schiemann J, Mendel RR, Salcheva G, Georgieva D. Stress and activity of molybdenum-containing complex (molybdenum co-factor) in winter wheat seeds. *Plant Physiol.* 1988; 87:533-535.

133. Wagner H, Michael G. Der Einfluss unterschiedlicher Stickstoffversorgung auf die Cytokinin bildung in Wurzeln von Sonnenblumen pflanzen. *Biochem Physiol. Pflanz.* 1971; 162:147-158.
134. Wahid A, Shabbir A. Induction of heat stress tolerance in barley seedlings by pre-sowing seed treatment with glycinebetaine. *Plant Growth Regul.* 2005; 46(2):133-141.
135. Wahid A, Gelani S, Ashraf M, Foolad MR. Heat tolerance in plants: an overview. *Environ. Exp. Bot.* 2007; 61:199-223.
136. Wahid A, Gelani S, Ashraf M, Foolad MR. Heat tolerance in plants: an overview. *Environ. Exp. Bot.* 2007; 61:199-223.
137. Walch-Liu P, Neumann G, Bangerth F, Engels C. Rapid effects of nitrogen form on leaf morphogenesis in tobacco. *J Exp. Bot.* 2000; 51:227-237.
138. Willenbrink J. Über Beziehungen zwischen proteinumsatz und Schwefelver-sorgung der chloroplast. *Z. Pflanzenphysiol.* 1967; 56:427-438.
139. Witt M, Barfield. Environmental stress and plant productivity. In *Handbook of Agricultural Productivity*, M. Recheigl JR. (Ed.). Boca Raton, FL: CRC Press, 1982, 347-374.
140. Wolff SP, Garner A, Dean RT. Free radicals, lipids, and protein breakdown. *Trends Biochem. Sci.* 1986; 11:27-31
141. Wyn Jones RG, Gorham J. Intra- and inter-cellular compartmentation of ions. In: Läuchli, A., Lüttge, U. (Eds.), *Salinity: Environment-Plants-Molecules*. Norwell, MA: Kluwer Academic Publishers, 2002, 159-180.
142. Yruela I. Copper in plants: acquisition, transport and interactions. *Funct. Plant Biol.* 2009; 36(5):409-430.
143. Zeid IM, Shedeed ZA. Response of alfalfa to putrescine treatment under drought stress, *Biol. Plant.* 2006; 50:635-640.
144. Zhang H, Jennings A, Barlow PW, Forde BG. Dual pathways for regulation of root branching by nitrate. *Proc. Natl. Acad. Sci. USA.* 1999; 96:6529-6534
145. Ziska LH, Tomecek MB, Gealy DR. Evaluation of competitive ability between cultivated and red weedy rice as a function of recent and projected increases in atmospheric CO₂. *Agron. J.* 2010; 102:118-123.